

# CHEMICAL CONSTITUENTS FROM SEEDS OF *Lactuca sativa*<sup>a</sup>

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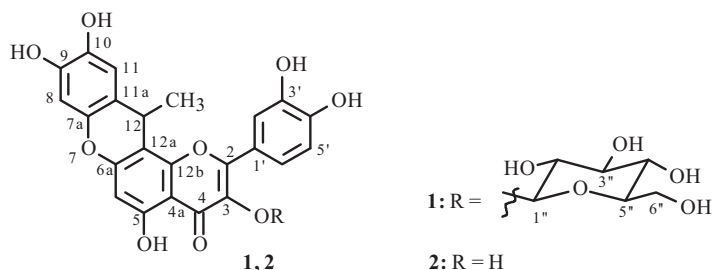
A new flavonol glycoside with a rare structure type, lactucasativoside A (**1**), was isolated from the seeds of *Lactuca sativa*, together with three known compounds, japonicin A (**2**), isoquercitrin (**3**), and caffeic acid (**4**). Their structures were established on the basis of extensive spectroscopic analysis. Compound **2** was obtained from the genus *Lactuca* for the first time.

**Keywords:** *Lactuca sativa* L., Compositae, flavonoids, lactucasativoside A.

*Lactuca sativa* L. (Compositae), a plant well known as lettuce, is a kind of favorite vegetable and is widely cultivated in China. The seeds of this plant are traditionally used as a folk medicine in China and Iran to relieve hyperpyrexia, inflammation, anuria, neurasthenia, gastrodynia, and osteodynia [1–3]. Phytochemical analysis showed the presence of triterpenoids, saponins, and phenols in the lettuce seeds [3].

In our recent research, the anti-inflammatory and analgesic activities of lettuce seeds were evaluated from mouse ear edema resulting from xylene and mouse writhing symptom caused by acetic acid. The results indicated that the 50% EtOH extract possessed the best anti-inflammatory activity, and the 95% EtOH extract showed the best analgesic activity. In order to reveal the active components, the chemical constituents of the seeds were isolated via extraction with 95% and 50% EtOH, purified by column chromatography and pre-HPLC, and identified on the basis of spectral data of MS and NMR. A new flavonol glycoside with a rare structure type, lactucasativoside A (**1**), together with three known compounds, japonicin A (**2**), isoquercitrin (**3**), and caffeic acid (**4**), was isolated from the seeds of *Lactuca sativa*.

Compound **1**, brown powder, had the molecular formula C<sub>29</sub>H<sub>26</sub>O<sub>14</sub>, determined by analysis of its (–) HR-ESI-MS (*m/z* 597.1238 [M – H]<sup>–</sup>, calcd for C<sub>29</sub>H<sub>25</sub>O<sub>14</sub> 597.1244), indicating the presence of 17 degrees of unsaturation. The <sup>1</sup>H and <sup>13</sup>C NMR data (Table 1) of **1** revealed structural features similar to those of japonicin A (**2**) [4], except for the presence of a sugar moiety. In addition, the chemical shift for C-2 was 9.8 ppm downfield (δ<sub>C</sub> 159.0) and C-3 was 1.9 ppm upfield (δ<sub>C</sub> 136.1). The glycosylation shifts indicated that the sugar unit was linked to C-3 of the aglycone, which was confirmed by the HMBC correlation of H-1'' (δ<sub>H</sub> 5.35) to C-3 (δ<sub>C</sub> 136.1). The remaining fragment connection was completely confirmed by HMBC correlations (Fig. 1).

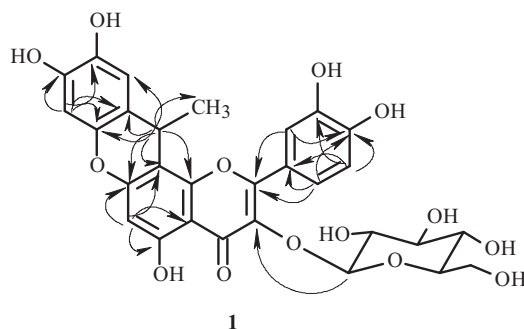


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TABLE 1.  $^1\text{H}$  NMR (600 MHz) and  $^{13}\text{C}$  NMR (150 MHz) Data of **1** and **2** ( $\text{CD}_3\text{OD}$ ,  $\delta$ , ppm, J/Hz)

| C atom | <b>1</b>                             |                     | <b>2</b>                    |                     |
|--------|--------------------------------------|---------------------|-----------------------------|---------------------|
|        | $\delta_{\text{H}}$                  | $\delta_{\text{C}}$ | $\delta_{\text{H}}$         | $\delta_{\text{C}}$ |
| 2      | —                                    | 159.0               | —                           | 149.2               |
| 3      | —                                    | 136.1               | —                           | 138.0               |
| 4      | —                                    | 179.7               | —                           | 177.7               |
| 4a     | —                                    | 108.2               | —                           | 107.1               |
| 5      | —                                    | 160.7               | —                           | 160.3               |
| 6      | 6.39 (1H, s)                         | 99.9                | 6.40 (1H, s)                | 99.3                |
| 6a     | —                                    | 158.4               | —                           | 158.1               |
| 7a     | —                                    | 144.3               | —                           | 144.6               |
| 8      | 6.53 (1H, s)                         | 104.4               | 6.57 (1H, s)                | 104.6               |
| 9      | —                                    | 146.1               | —                           | 146.2               |
| 10     | —                                    | 143.5               | —                           | 143.6               |
| 11     | 6.75 (1H, s)                         | 115.1               | 6.80 (1H, s)                | 115.3               |
| 11a    | —                                    | 117.1               | —                           | 116.0               |
| 12     | 4.35 (1H, q, J = 7.2)                | 28.7                | 4.43 (1H, q, J = 7.2)       | 28.9                |
| 12a    | —                                    | 106.4               | —                           | 106.4               |
| 12b    | —                                    | 154.7               | —                           | 154.5               |
| 1'     | —                                    | 123.1               | —                           | 124.3               |
| 2'     | 7.81 (1H, d, J = 1.8)                | 117.4               | 7.86 (1H, d, J = 1.8)       | 117.4               |
| 3'     | —                                    | 146.1               | —                           | 146.6               |
| 4'     | —                                    | 150.2               | —                           | 148.5               |
| 5'     | 6.93 (1H, d, J = 8.4)                | 116.2               | 6.97 (1H, d, J = 8.4)       | 116.6               |
| 6'     | 7.72 (1H, dd, J = 8.4, 1.8)          | 123.3               | 7.76 (1H, dd, J = 8.4, 1.8) | 121.9               |
| 12-Me  | 1.44 (3H, d, J = 7.2)                | 26.3                | 1.51 (3H, d, J = 7.2)       | 26.3                |
| 1''    | 5.35 (1H, d, J = 7.8)                | 104.1               |                             |                     |
| 2''    | 3.52 (1H, t, J = 7.8)                | 75.7                |                             |                     |
| 3''    | 3.45 (1H, t, J = 7.8)                | 78.1                |                             |                     |
| 4''    | 3.37 (1H, t, J = 7.8)                | 71.3                |                             |                     |
| 5''    | 3.25 (1H, m)                         | 78.5                |                             |                     |
| 6''    | 3.73 (1H, dd, J = 12.0, 2.4, H-6''a) | 62.5                |                             |                     |
|        | 3.59 (1H, dd, J = 12.0, 5.4, H-6''b) |                     |                             |                     |

Fig. 1. Key HMBC correlations of compound **1**.

Acid hydrolysis of **1** afforded  $\beta$ -D-glucose as confirmed by comparison with an authentic sample and established by the J value ( $J = 7.8$  Hz) of the anomeric proton (H-1''). Thus, compound **1** was readily established as 3,3',4',5,9,10-hexahydroxy-12-methylchroman[2,3-h]flavone 3-O- $\beta$ -D-glucopyranoside, named lactucasativoside A (**1**).

Compounds **2–4** were identified as japonicin A (**2**) [4], isoquercitrin (**3**) [5], and caffeic acid (**4**) [5], respectively, by comparing their spectroscopic data with those reported in the literature and were isolated from the seeds of *Lactuca sativa* for the first time.

## EXPERIMENTAL

**General Experimental Procedures.** Melting points were determined on a Yanaco MP-S3 apparatus (uncorrected). Optical rotation was measured on a Rudolph Autopol IV polarimeter. UV spectroscopy was carried out on a Shimadzu UV-2550 spectrophotometer. IR spectrum was obtained using a Bio-Rad FTS 165 spectrophotometer. NMR spectra were recorded on a Varian VNMRS 600 spectrometer. HR-ESI-MS data were acquired using a Q STAR Elite mass spectrometer (Applied Biosystems). D101 macroporous resin (Cangzhou Baoen Chemical Co., Ltd, China), silica gel (Qingdao Haiyang Chemical Co., Ltd, China), and Sephadex LH-20 (Amersham Pharmacia Biotech, Sweden) were used for chromatography.

**Plant Material.** The seeds of *Lactuca sativa* L. were collected from the Miquan Vegetable Planting Base in Xinjiang, China in 2008. The seeds were authenticated by chief researcher Y. F. Zhang at the Xinjiang Institute of Materia Medica. A voucher specimen was deposited in the Herbarium of the Xinjiang Institute of Materia Medica.

**Extraction and Isolation.** The dried seeds (10 kg) were extracted with 95% EtOH and 50% EtOH (2 × 100 L) successively to give a crude extract (1108 g) after evaporation *in vacuo*. The extract was suspended in water and extracted with petroleum ether. The aqueous layer (614 g) was condensed and chromatographed over a macroporous absorbent resin column with gradient elution by water, 30% EtOH, 50% EtOH, 70% EtOH, and 95% EtOH successively. The 50% EtOH fraction (15 g) was chromatographed over a silica gel column, eluted with CHCl<sub>3</sub>–CH<sub>3</sub>OH (50:1–2:8) in a gradient manner to give six fractions. Fraction 3 was subjected to Sephadex LH-20 (eluted with MeOH) to give 13 subfractions. Fraction 3–9 was purified by prep-HPLC using CH<sub>3</sub>OH–0.2% HCOOH (45:55) as eluent to afford **1** (10 mg). Fraction 3–12 was purified through Sephadex LH-20 (MeOH) to give **2** (3 mg). Fraction 3–8 was purified by Sephadex LH-20 (MeOH) to give **3** (15 mg). The 30% EtOH fraction (55 g) was subjected to silica gel column chromatography with gradient elution by CHCl<sub>3</sub>–CH<sub>3</sub>OH (50:1–2:8) to give nine fractions. Fraction 7 was chromatographed over Sephadex LH-20 gel (MeOH) to yield **4** (5 mg).

**Lactucasativoside A (1).** C<sub>29</sub>H<sub>26</sub>O<sub>14</sub>, brown powder (MeOH), mp > 270°C.  $[\alpha]_D^{20} -10.00^\circ$  (*c* 0.01, MeOH). UV (MeOH,  $\lambda_{\max}$ , nm) (log  $\epsilon$ ): 227 (3.98), 250 (3.74), 270 (3.76), 274 (3.84), 355 (3.66). IR (KBr,  $\nu$ , cm<sup>-1</sup>): 3368, 1656, 1597, 1518, 1488, 1456, 1383, 1351, 1261, 1180, 1071. (–) HR-ESI-MS *m/z* 597.1238 [M – H]<sup>–</sup> (calcd for C<sub>29</sub>H<sub>25</sub>O<sub>14</sub>, 597.1244). <sup>1</sup>H and <sup>13</sup>C NMR (Table 1).

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